

Calcium Phosphate Bone Cements in Drug Delivery: Review

İlaç Verilişinde Kalsiyum Fosfat Kemik Çimentoları

Evren ALĞİN YAPAR^a

^aRepublic of Turkey, Ministry of Health,
Turkish Medicines and Medical Devices
Agency, Ankara

Geliş Tarihi/Received: 09.10.2013
Kabul Tarihi/Accepted: 06.11.2013

Yazışma Adresi/Correspondence:
Evren ALĞİN YAPAR
Republic of Turkey, Ministry of Health,
Turkish Medicines and
Medical Devices Agency, Ankara,
TÜRKİYE/TURKEY
evrenalgin@yahoo.com

ABSTRACT In recent years, local drug administration has gain importance due to increase in the success in the field of orthopedic surgery. Accordingly implants has been prepared in various forms with different materials. Researches have been carried out regarding to their *in vitro* and *in vivo* performances. Between orthopedic implant applications, cement forms have been found clinically successful and calcium phosphate cements with their similarity to bone structure have gain attention in drug delivery due to their advantages such as injectability and fast setting time in low-temperatures providing that their mechanical properties develop by combination of other biomaterials. Several attempts have been made to include growth factors and morphogens within bioactive scaffolds to stimulate cellular adhesion, proliferation and differentiation, so as to promote bone regeneration. Additionally, enhancing further the functionality of these already complex cements by loading drugs into them to treat bone disorders or to act on the surrounding tissues with an adequate therapeutic concentration level and for a desired time frame is recognized as being highly beneficial. Successful researches are available with especially to antibiotics, biphosphanates, growth factors, anti-inflammatory and antimicrobial actives in the form of calcium phosphate cements and this field is promising for new applications of the future for musculoskeletal diseases and defects.

Key Words: Drug delivery systems; biocompatible materials; bone cements; calcium phosphate; orthopedics

ÖZET Son yıllarda ortopedi alanında lokal ilaç verilışı, özellikle cerrahi mühalelerin başarısını arttırması nedeniyle önem kazanmıştır. Bu doğrultuda çeşitli materyaller ile değişik formlarda implantlar hazırlanmış, *in vitro* ve *in vivo* performanslarına yönelik araştırmalar yapılmıştır. Ortopedik implant uygulamaları arasında, klinik olarak çimento formları başarılı bulunmuş ve materyal olarak kemik yapısı ile oldukça benzer kalsiyum fosfatlar, mekanik özelliklerinin diğer biyomateriyaller ile geliştirilmesi kaydıyla ilaç salınında enjekte edilebilirlik, düşük sıcaklıkta hızlı sertleşme gibi avantajları nedeniyle dikkat çekmiştir. Kemik rejenerasyonu teşvik etmek amacıyla, hücre yapışması, çoğalması ve farklılaşması, uyarılmak için biyolojik olarak aktif iskeleler içinde büyüme faktörleri ve morfojenlerin verilmesi amacıyla çeşitli girişimler yapılmıştır. İlaveten, kemik bozukluklarını tedavi etmek veya çevre dokularda etki etmek üzere yeterli terapötik konsantrasyon seviyesinin arzu edilen zaman dilimi içinde sağlanması amacıyla bu kompleks çimentolara ilaç yüklenerek işlevselliğinin artırılması son derece faydalı olarak kabul edilmektedir. Başta antibiyotikler olmak üzere bifosfonatlar, büyüme faktörleri, antiinflatuarlar ve antimikrobiallerin kalsiyum fosfat kemik çimentosu formunda başarılı çalışmaları mevcut olup, ileriye yönelik kas ve iskelet sistemi hastalık ve defektlerinde yeni uygulamalar açısından umut vericidir.

Anahtar Kelimeler: İlaç dağıtım sistemleri; biyoyoumlu materyaller; kemik çimentosu; kalsiyum fosfat; ortopedi

Türkiye Klinikleri J Pharm Sci 2013;2(1):34-41

In recent years drug delivery implants have gain importance for long term therapies which have some advantages like avoidance of first pass metabolism, reduce dose and side effects of active substance with fluctuations in blood levels.¹

In orthopedic surgery osteoclasts, one of the major causes of bone lysis leading to implant failure has been clinically proved that could be effectively sponge out by decreasing the catabolic bone activity provided with bisphosphonate group drugs. Due to systemic delivery of bisphosphonate by injections could not effectively reach to peri-implant region; local drug delivery has been preferred.² Drug delivery orthopedic implants particularly focus on bone infections which is an important factor in the success of orthopedic implants. Drug delivery with an implant can be available either coated on an implant surface, or incorporated in a biomaterials/cement scaffold or included in beads (Figure 1). Between those approaches most of clinical orthopedic applications were performed in cements successfully.²⁻⁵

Several attempts have been made to include growth factors and morphogens within bioactive scaffolds to stimulate cellular adhesion, proliferation and differentiation, so as to promote bone regeneration.⁶⁻⁸ In addition, enhancing further the functionality of these already complex matrices by loading drugs into them to treat bone disorders or to act on the surrounding tissues with an adequate therapeutic concentration level and for a desired time frame is recognized as being highly beneficial.^{6,9-20}

Three-dimensional bioactive bone scaffolds can be fabricated by using bioceramics, biodegradable polymers and their composites. Bioactive ce-

ramic scaffolds alone used in bone tissue engineering can serve as a delivery vehicle for drugs but the drug release patterns are difficult to control. On the other hand, biodegradable polymeric materials such as poly(lactic-co-glycolic acid) (PLGA) and poly(propylene glycol-fumerate)/methylmethacrylate can be used to control the local delivery of drugs. However, they can show impaired osteoconduction and they can provoke an adverse tissue response owing to inflammation as a consequence of acidic degradation.^{6,7,21-24} Inorganic materials are promising alternatives to polymeric bone cements and fillers because of their high chemical stability, hydrophilic character, easy functionalization, large surface area and ability to adsorb drugs. Owing to the possibility of synthesizing ordered mesoporous silica-based structures, inorganic particulate fillers can achieve a controlled release of the adsorbed or attached moiety. However self-supported structures of those materials lack mechanical stability.^{25,26} Composite materials might combine for improvement of mechanical properties and adjustable controlled release of drug which can be obtained with combination of polymeric and inorganic (hydroxyapatites, tricalcium phosphates and mesoporous silicas) materials. Mechanically, a well-designed composite material could simulate at best the behavior of natural bone, which is composed of an inorganic (calcium hydroxyapatite) and an organic (type I collagen and other non-collagenous proteins) matrix.²⁶ Thus, the smart combination of bioceramics and biodegradable polymers can not only improve the degradability of the inorganic material and alter its mechanical/physical properties, but also drug-release profiles can be controlled to a greater extent rather than pure ceramics. There is a wide range of different polymers that can be use for such applications, having different degradation rates and mechanisms, and a wide range of bioceramic/biopolymer composite scaffolds is available for bone tissue engineering.^{6,7,21,26-29}

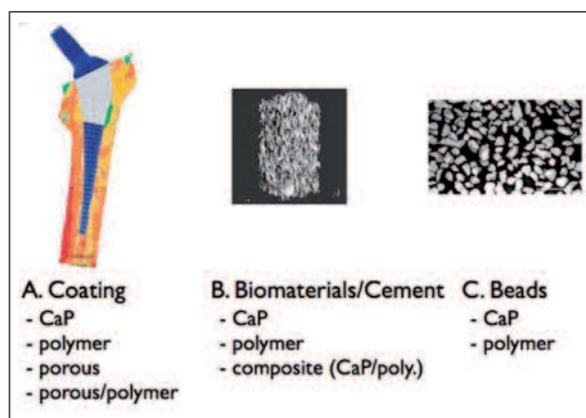


FIGURE 1: Drug delivery implant approaches A. Coating, B. Scaffold or Cement, C. Beads.²

(See color figure at

<http://www.turkiyeklinikleri.com/journal/eczacilik-bilimleri-dergisi/2146-944X/>)

CALCIUM PHOSPHATE CEMENTS

GENERAL PROPERTIES

Calcium phosphates (CaPs) have good biocompatibility, osteoconductivity and chemical properties

make them suitable for bone-remodeling kinetics and they can be resorbed by cells. They have enough mechanical strength but they are brittle and slowly degradable.^{6,30-32} Their osteoconductive properties allowed them to use in orthopedics, dental, ear, nose and throat surgeries.³³ CaPs can be in different forms such as ceramics, cements, composites and thin coatings.³³⁻³⁵ Their performance can be affected by some pathological situations such as infection, irradiation, diseases etc. in terms of the substitution and/or resorption process. To prevent the undesirable results related pathological situations, CaPs preferred to combine with active molecules which represent the most current alternatives to biological bone grafts and exist in various forms such as powders, granules, ceramic, cement and coatings.³³ On the basis of composition, synthetic CaPs presently used as biomaterials are classified as presented in Table 1.

Drug delivery bone cements should be bioactive and resorbable to provide bonding to bone tissue and substitution. Moreover injectability is desirable for ease of administration and better fitting to the bone defect. Calcium phosphate cements (CPCs) meet above mentioned needs which make them good candidates for clinical applications.³⁷ The CPCs was first patent by Brown and Chow in 1986.^{33,38} CPCs have more advantages such

as being noncytotoxic, promoting the development of osteoconductive pathways, enough mechanical strength for different applications and restore contour. Furthermore, their low-temperature setting reaction and intrinsic porosity allow for the incorporation of drugs into the cement. Fast setting time, excellent mouldability, easy manipulation and injectability can perfectly fit the bone defect; their low temperature *in vivo* self-setting capability is important not to give harm to surrounding tissue and provide no risk of infectious diseases allow to prevent grafting failure.^{33,37,39-46}

CPCs are composed of an aqua/aqueous solution and CaPs or combinations. Upon mixing of these components, dissolution and precipitation into a less soluble CaP has occurred and entanglement of the growth of crystals provided the mechanical rigidity of the cement.^{36,39,46} The prepared CaPs paste can be placed into the damaged part of bone and generally hardens less than 20 min at body temperature (37°C) *in situ* and then displays limited solubility. Stability and solubility of CaPs and their combinations is the major collimator for the setting process which also dependent upon the pH value of a cement paste.⁴⁶ Various CPCs are currently commercially and many more are in experimental stages.³³

MECHANICAL PROPERTIES

Although having many advantages, CPCs have limitations owing to their poor mechanical properties and slow *in vivo* biodegradation.³⁶ Mechanical properties of CPCs such as strength, setting time, porosity and swelling can be controlled by liquid-to-powder ratio, pH of the liquid phase, powder composition, chemistry, crystallinity, particle size, presence of nucleating agents in the reaction system are other important factors for mechanical properties.^{36,47-52}

The current commercial CPCs remain dense after implantation and insufficient macroporosity will take advantage of 3D cell colonization and tissue ingrowth.³³ In this context, recently CPCs have been designed with polysaccharides or resorbable fibers which are supposed to develop channels suitable for bone ingrowth by dissolution of these particles or fibers.^{33,53-55}

TABLE 1: Calcium phosphate types, abbreviations and mineral names.³⁶

Calcium phosphates	Abbreviations	Mineral name
Monocalcium phosphate monohydrate	MCPM	–
Monocalcium phosphate	MCP	–
Dicalcium phosphate dihydrate	DCPD	Brushite
Dicalcium phosphate	DCP	Monetite
Octacalcium phosphate	OCP	–
alpha-Tricalcium phosphate	α-TCP	–
beta-Tricalcium phosphate	β-TCP	Whitelockite
Amorphous calcium phosphate	ACP	–
Calcium-deficient hydroxyapatite	CDHA	–
Carbonated apatite	CA	Dahlite
Hydroxyapatite	HA	–
Oxyapatite	OXA	–
Tetracalcium phosphate	TTCP	Hilgenstockite

Polymers are usually added to CPC to increase the mechanical properties and control degradation.^{36,56-58} Increased setting time and reduced workability are also reported with increased mechanical properties.^{36,59-61} Polymeric materials such as chitosan,^{62,63} alginate,^{64,65} gelatin,⁶⁶⁻⁶⁸ poly(acrylic acid),⁶⁹ polymethylmethacrylate,⁷⁰ PLGA,⁷¹ pectin⁷² have been used to improve the anti-washout and handling properties of CPCs as these materials tend to disintegrate on early contact with blood and other fluid.³⁶ The undesired effects of organic additives reported as delay in setting time and decrease in mechanical strength.⁵¹ The setting reaction of CPCs can be affected or modified by adding an active molecule to the powder phase or the liquid phase which could be resulted as a change in physico-chemical and mechanical properties.^{37,69,73-77} In general, in apatitic cements, antibiotics have a tendency to increase setting times and reduce the mechanical strength of the cements.^{37,73-76} This decrease of mechanical strength can be attributed to different factors, such as increased porosity or to some inhibition of the setting reaction, as suggested by the presence of certain amount of reactants in the set cements when the antibiotic quantity increases. In other cases the change in the setting properties are caused by some chemical interaction with the drug, which can modify the kinetics of the dissolution-precipitation reaction and the morphology of the precipitated crystals.³⁷ The highly microporous structure of CPC can be obtained different liquid-to-powder ratios, after setting, allows it to incorporate drugs into its structure. More compacted cement microstructure with smaller size of pores can be obtained by decreasing the liquid-to-powder ratio. The drug can be introduced either in the liquid or the solid phase of the CPC, but the physicochemical properties of the drug or protein must be considered for do not change during the chemical reaction and setting of CPC.^{36,52,78} Studies particularly for antibiotics showed the relation between drug concentration and CPCs structure by means of incorporated drug effects on the structure of CPCs.^{36,79-81} A morphological change and decrease in compressive strength of CPC with in-

creases in tetracycline concentration was reported and referred as the strong affinity of tetracycline hydrochloride and addressed this limitation to some extent by treating tetracycline hydrochloride with CaP solution and then incorporating it into CPC.^{36,80} A maximum of 7% cephalixin monohydrate was incorporated without affecting the mechanical properties of CPC and also observed an increase in setting time and decrease in crystallinity of CPC.⁷⁹ An increase in setting time was also reported due to gentamicin sulphate incorporation into the CPC matrix.⁸¹ Drug release from CPC depends also on the intrinsic porosity, which is consequence of processing parameters.^{36,78,81} Despite excellent osteoconductivity and good applicability, CPCs use in drug delivery is limited which is mostly due to the changes in the final properties of CPCs resulting from the drug incorporation, changes in the drug activity and its bioavailability.³⁶

CPCs are available to use both as bone substitutes and drug carriers for treatments of different skeletal diseases and bone fracture healings. The drugs can be incorporated throughout the whole material volume of CPCs without losing activity and denaturalization by adding them into one of the two cement phases which can facilitate the release of drugs for more prolonged times. Several studies related to the application of both commercial and experimental CPCs as drug carriers for local or systemic treatments for different durations have been published.³⁷

DRUG RELEASE BEHAVIORS

Release behaviors from CPCs are influenced by physico-chemical properties such as drug solubility and chemical property, microstructure, crystallinity, density and porosity of the final CPC. Changes during hydration and setting are also effective on drug-cement interactions, and degradation behavior of CPCs. If a polymer is used in the CPC matrix, drug release kinetics also depends on its solubility, molecular weight, drug-polymer interactions and degradation rate. Generally, depending on the drug release behavior, drug

delivery devices can be categorized mainly as diffusion controlled and activation controlled systems which also can be defined as i. diffusion controlled ii. chemical processes controlled, and iii. externally or electronically controlled.^{1,36,41} The degradation of drug incorporated CPC matrix is usually a slower process than the drug release kinetics, thus release kinetics is generally a diffusion dominated process from the biodegradable CPCs. Degradation related release is also a simultaneous process with this diffusion.^{36,82} Figure 2 illustrates the schematic of drug release from a CPC loaded with drug molecules.³⁶ Diffusion dominated release kinetics from a matrix can be described by the square root of time kinetics namely Higuchi law which is based on Fickian diffusion under the assumption that drug molecules are uniformly dispersed in a homogeneous matrix. For longer duration, release kinetics does not always follow the Higuchi law due to other factors such as changes in cement matrix composition.^{36,69,82,83} Hydrophobic-hydrophilic interactions between drug-polymer and drug-release medium also could influence the release kinetics, which might not follow simple power laws.³⁶

TREATMENT APPROACHES FOR CPCs

Biocompatibility and nonexothermic behavior of CPCs are important factors in drug incorporating attempts and CPC used for antibiotic delivery gives good clinical results^{33,39,84-87} except some resistance strains. Avoidance of the routine use of such drug loaded cements and restricting their use only to multiresistant strains are recommended by some clinicians.^{33,88-90} On the other hand further studies clearly showed that CaP matrices are good carriers for controlling the catabolic bone remodeling drugs.^{2,91-93}

CPCs can be used for local drug delivery for the treatment of different skeletal diseases such as bone tumors, osteoporosis or osteomyelitis. The types of drug-eluting implants used in traumatology and in orthopedic surgery include: **i)** antibiotic-loaded bone cements and fillers used to prevent infection (osteomyelitis) in orthopedic surgery; **ii)** cements

loaded with osteoinductive molecules such as growth factors to favor the osseointegration of the implant; and **iii)** devices loaded with chemotherapeutic agents, antiestrogens or anti-inflammatories used to treat different pathologies including osteosarcomas and degenerative diseases.²⁶

In this context, many kinds of drugs/active molecules, including antibiotics,^{3,79,94-96} chemotherapeutics,^{36,97-99} growth factors,^{4,36} proteins/amino acids,^{36,100,101} antimicrobial peptides (AMPs),^{36,102,103} nonsteroidal analgesic and anti-inflammatory drugs (NSAIDs)¹⁰⁴ and bisphosphonates^{90,105,106} have been incorporated into CPCs for various applications. Table 2 refers some research studies on CPCs with different drugs published in last decades.

CONCLUSION

As a conclusion development of new dosage forms by using biomaterials is important in drug delivery since it is hard to develop new drug molecules. In this respect drug delivery either local or systemic via bone cements, particularly with CPCs owing to their biocompatibility, noncytotoxicity, osteoconductivity, low-temperature setting reaction and fast setting time, mouldability, easy manipulation and injectibility would make them good alternative and beneficial application in musculoskeletal diseases and defects.

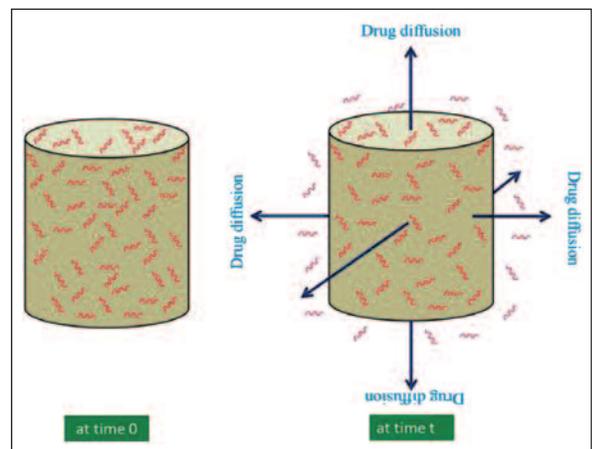


FIGURE 2: Drug delivery implant approaches A. Coating, B. Scaffold or Cement, C. Beads.²

(See color figure at

<http://www.turkiyeklinikleri.com/journal/eczacilik-bilimleri-dergisi/2146-944X/>)

TABLE 2: Some research studies on CPCs with different drugs published between 2003 and 2013 years.

Cement Composition	Active ingredient	Group	References
Silica-CaP	Gentamicin	Antibiotic	3
HA	Bone marrow stroma cell	Stem cell	31
α -TCP,DCP,CaCO ₃ , HA,PLGA	rhBMP-2	Growth factor	71
DCPD	Vancomycin	Antibiotic	82
Sr-substituted β -TCP	Doxycycline hyclate	Antibiotic	83
HA	Zoledronate	Bisphosphonate	91
HA:calcium sulphate mixture	Cephalexin	Antibiotic	96
α -TCP, DCP, HA	Paclitaxel	Chemotherapeutic	99
TTCP:DCPA mixture and Chitosan	rhBMP-2	Protein	100
TTCP, DCP	Gentamicin	Antibiotic	103
β -TCP	Ibuprofen	NSAIDs	104
α -TCP	Alendronate	Bisphosphonate	106
TTCP:DCPA mixture and Chitosan	human bone marrow mesenchymal stem cell (hBMSC)	Protein	107
ACP, DCPD	rhBMP-2	Growth factor	108
Sr- substituted HA	Gentamicin	Antibiotic	109
α -TCP, alginate	Rat bone mesenchymal stem cells	Protein	110
nano-hydroxyapatite/chitosan (n-HA/CS)	Berberine	Antimicrobial	111
HA	Tetracycline hydrochloride	Antibiotic	112
DCPD and TTCP	Zoledronic acid	Bisphosphonate	113

REFERENCES

- Alğın Yapar E. [Overview on implantable polymeric drug delivery systems: Review]. *Türkiye Klinikleri J Pharm Sci* 2012;1(2):73-82.
- Pioletti DP, Gauthier O, Stadelmann VA, Bujoli B, Guicheux J, Zambelli PY, et al. Orthopedic implant used as drug delivery system: clinical situation and state of the research. *Curr Drug Deliv* 2008;5(1):59-63.
- El-Ghannam A, Ahmed K, Omran M. Nanoporous delivery system to treat osteomyelitis and regenerate bone: gentamicin release kinetics and bactericidal effect. *J Biomed Mater Res B Appl Biomater* 2005;73(2):277-84.
- Blom EJ, Klein-Nulend J, Wolke JG, van Waas MA, Driessens FC, Burger EH. Transforming growth factor-beta1 incorporation in a calcium phosphate bone cement: material properties and release characteristics. *J Biomed Mater Res* 2002;59(2):265-72.
- Denissen H, van Beek E, van den Bos T, de Bleeck J, Klein C, van den Hooff A. Degradable bisphosphonate-alkaline phosphatase-complexed hydroxyapatite implants in vitro. *J Bone Miner Res* 1997;12(2):290-7.
- Mouriño V, Boccaccini AR. Bone tissue engineering therapeutics: controlled drug delivery in three-dimensional scaffolds. *J R Soc Interface* 2010; 7(43):209-27.
- Habraken WJ, Wolke JG, Jansen JA. Ceramic composites as matrices and scaffolds for drug delivery in tissue engineering. *Adv Drug Deliv Rev* 2007;59(4-5):234-48.
- Lee SH, Shin H. Matrices and scaffolds for delivery of bioactive molecules in bone and cartilage tissue engineering. *Adv Drug Deliv Rev* 2007; 59(4-5):339-59.
- Gomes ME, Reis RL. Biodegradable polymers and composites in biomedical applications from catgut to tissue engineering. Part II. Systems for temporary replacement and advanced tissue regeneration. *Int Mater Rev* 2004;49(5):274-85.
- Duarte AR, Mano JF, Reis RL. Dexamethasone-loaded scaffolds prepared by supercritical-assisted phase inversion. *Acta Biomater* 2009;5(6): 2054-62.
- Baroli B. From natural bone grafts to tissue engineering therapeutics: Brainstorming on pharmaceutical formulative requirements and challenges. *J Pharm Sci* 2009;98(4):1317-75.
- Sayın B, Çaliş S. [Bone grafts: Review]. *TÜFTAD Haberler* 2006;13(4):3-7.
- Gürsel I, Korkusuz F, Türesin F, Alaeddinoglu NG, Hasirci V. In vivo application of biodegradable controlled antibiotic release systems for the treatment of implant-related osteomyelitis. *Biomaterials* 2001;22(1):73-80.
- Samdancıoğlu S, Calis S, Sumnu M, Atilla Hincal A. Formulation and in vitro evaluation of bisphosphonate loaded microspheres for implantation in osteolysis. *Drug Dev Ind Pharm* 2006;32(4): 473-81.
- Şamdancıoğlu S, Çaliş S, Şumnu M, Hincal AA. In vitro studies on alendronate sodium incorporated biodegradable alginate beads for implantation. *FABAD J Pharm Sci* 2005;30(3): 142-9.
- Orhan Z, Cevher E, Mülazımoğlu L, Gürcan D, Alper M, Araman A, et al. The preparation of ciprofloxacin hydrochloride-loaded chitosan and pectin microspheres: their evaluation in an animal osteomyelitis model. *J Bone Joint Surg Br* 2006;88(2):270-5.
- Cevher E, Orhan Z, Mülazımoğlu L, Sensoy D, Alper M, Yıldız A, et al. Characterization of biodegradable chitosan microspheres containing vancomycin and treatment of experimental osteomyelitis caused by methicillin-resistant *Staphylococcus aureus* with prepared microspheres. *Int J Pharm* 2006;317(2):127-35.
- Orhan Z, Cevher E, Yıldız A, Ahiskali R, Sensoy D, Mülazımoğlu L. Biodegradable microspherical implants containing teicoplanin for the treatment of methicillin-resistant *Staphylococcus aureus* osteomyelitis. *Arch Orthop Trauma Surg* 2011;130(1): 135-42.
- Ozcan I, Bouchemal K, Segura-Sánchez F, Ozer O, Güneri T, Ponchel G. Synthesis and characterization of surface-modified PBLG nanoparticles for bone targeting: in vitro and in vivo evaluations. *J Pharm Sci* 2011;100(11): 4877-87.
- Ozcan I, Segura-Sánchez F, Bouchemal K, Sezak M, Ozer O, Güneri T, et al. Pegylation of poly(γ -benzyl-L-glutamate) nanoparticles is efficient for avoiding mononuclear phagocyte system capture in rats. *Int J Nanomedicine* 2010;5:1103-11.

21. Rezwani K, Chen QZ, Blaker JJ, Boccaccini AR. Biodegradable and bioactive porous polymer/inorganic composite scaffolds for bone tissue engineering. *Biomaterials* 2006;27(18): 3413-31.
22. Garvin K, Feschuk C. Polylactide-polyglycolide antibiotic implants. *Clin Orthop Relat Res* 2005; 437:105-10.
23. Gerhart TN, Roux RD, Hanff PA, Horowitz GL, Renshaw AA, Hayes WC. Antibiotic-loaded biodegradable bone cement for prophylaxis and treatment of experimental osteomyelitis in rats. *J Orthop Res* 1993;11(2):250-5.
24. Böstman O, Pihlajamäki H. Clinical biocompatibility of biodegradable orthopaedic implants for internal fixation: a review. *Biomaterials* 2000;21(24): 2615-21.
25. Wang S. Ordered mesoporous materials for drug delivery. *Microp Mesop Mater* 2009;117 (1-2):1-9.
26. Arruebo M, Vilaboa N, Santamaria J. Drug delivery from internally implanted biomedical devices used in traumatology and in orthopedic surgery. *Expert Opin Drug Deliv* 2010;7(5): 589-603.
27. Nair LS, Laurencin CT. Biodegradable polymers as biomaterials. *Prog Polym Sci* 2007;32(8-9): 762-8.
28. Guarino V, Causa F, Ambrosio L. Bioactive scaffolds for bone and ligament tissue. *Expert Rev Med Devices* 2007;4(3):405-18.
29. Yunos DM, Bretcanu O, Boccaccini AR. Polymer-bioceramic composites for tissue engineering scaffolds. *J Mater Sci* 2008;43(13): 4433-42.
30. Karageorgiou V, Kaplan D. Porosity of 3D biomaterial scaffolds and osteogenesis. *Biomaterials* 2005;26(27):5474-91.
31. Marcacci M, Kon E, Moukhachev V, Lavroukov A, Kutepov S, Quarto R, et al. Stem cells associated with macroporous bioceramics for long bone repair: 6- to 7-year outcome of a pilot clinical study. *Tissue Eng* 2007;13(5):947-55.
32. Bretcanu O, Chen QZ, Boccaccini AR. Inorganic and composite bioactive scaffolds for bone tissue engineering. In: Liu X, Chu PK, eds. *Biomaterials Fabrication And Processing Handbook*. 1st ed. Boca Raton, FL: CRC Press; 2008. p.3-43.
33. Verron E, Khairoun I, Guicheux J, Boulter JM. Calcium phosphate biomaterials as bone drug delivery systems: a review. *Drug Discov Today* 2010;15(13-14):547-52.
34. Rush SM. Bone graft substitutes: osteobiologics. *Clin Podiatr Med Surg* 2005;22(4):619-30, viii.
35. Vallet-Regí M. Revisiting ceramics for medical applications. *Dalton Trans* 2006;44:5211-20.
36. Bose S, Tarafder S. Calcium phosphate ceramic systems in growth factor and drug delivery for bone tissue engineering: a review. *Acta Biomater* 2012;8(4):1401-21.
37. Ginebra MP, Canal C, Espanol M, Pastorino D, Montufar EB. Calcium phosphate cements as drug delivery materials. *Adv Drug Deliv Rev* 2012;64(12):1090-110.
38. Brown WE, Chow LC. Effects of neutral salts in a bench-scale caries model. *J Dent Res* 1986; 65(9):1115-20.
39. Ginebra MP, Traykova T, Planell JA. Calcium phosphate cements as bone drug delivery systems: a review. *J Control Release* 2006;113(2): 102-10.
40. Bohner M. Reactivity of calcium phosphate cements. *J Mater Chem* 2007;17(38):3980-6.
41. Ginebra MP, Traykova T, Planell JA. Calcium phosphate cements: competitive drug carriers for the musculoskeletal system? *Biomaterials* 2006; 27(10):2171-7.
42. Bohner M. Calcium orthophosphates in medicine: from ceramics to calcium phosphate cements. *Injury* 2000;31(Suppl 4):37-47.
43. Dorozhkin SV. Calcium orthophosphate cements for biomedical application. *J Mater Sci* 2008;43(9):3028-57.
44. Sugawara A, Fujikawa K, Takagi S, Chow LC, Nishiyama M, Murai S. Histopathological and cell enzyme studies of calcium phosphate cements. *Dent Mater J* 2004;23(4):613-20.
45. Smartt JM Jr, Karmacharya J, Gannon FH, Ong G, Jackson O, Bartlett SP, et al. Repair of the immature and mature craniofacial skeleton with a carbonated calcium phosphate cement: assessment of biocompatibility, osteoconductivity, and remodeling capacity. *Plast Reconstr Surg* 2005;115(6):1642-50.
46. Pina S, Ferreira JMF. Brushite-forming Mg-, Zn- and Sr-substituted bone cements for clinical applications. *Materials* 2010;3(1):519-35.
47. Bohner M, Brunner TJ, Stark WJ. Controlling the reactivity of calcium phosphate cements. *J Mater Chem* 2008;18(46):5669-75.
48. Brunner TJ, Grass RN, Bohner M, Stark WJ. Effect of particle size, crystal phase and crystallinity on the reactivity of tricalcium phosphate cements for bone reconstruction. *J Mater Chem* 2007;17(38):4072-8.
49. Fernandez E, Boltong MG, Ginebra MP, Driessens FCM, Bermudez O, Planell JA. Development of a method to measure the period of swelling of calcium phosphate cements. *J Mater Sci Lett* 1996;15(11):1004-5.
50. Ginebra MP, Driessens FC, Planell JA. Effect of the particle size on the micro and nanostructural features of a calcium phosphate cement: a kinetic analysis. *Biomaterials* 2004;25(17):3453-62.
51. Ginebra MP, Boltong MG, Fernandez E, Planell JA, Driessens FCM. Effect of various additives and temperature on some properties of an apatitic calcium phosphate cement. *J Mater Sci: Mater Med* 1995;6(10):612-6.
52. Shahrouzi J, Hesaraki S, Zamanian A. The effect of paste concentration on mechanical and setting properties of calcium phosphate bone cements. *Adv Chem Eng Res* 2012;1(1): 1-7.
53. Xu HH, Takagi S, Quinn JB, Chow LC. Fast-setting calcium phosphate scaffolds with tailored macropore formation rates for bone regeneration. *J Biomed Mater Res A* 2004; 68(4):725-34.
54. Weir MD, Xu HH. High-strength, in situ-setting calcium phosphate composite with protein release. *J Biomed Mater Res A* 2008;85(2):388-96.
55. Xu HH, Quinn JB. Calcium phosphate cement containing resorbable fibers for short-term reinforcement and macroporosity. *Biomaterials* 2002;23(1):193-202.
56. Durucan C, Brown PW. Calcium-deficient hydroxyapatite-PLGA composites: mechanical and microstructural investigation. *J Biomed Mater Res* 2000;51(4):726-34.
57. Durucan C, Brown PW. Low temperature formation of calcium-deficient hydroxyapatite-PLA/PLGA composites. *J Biomed Mater Res* 2000;51(4):717-25.
58. Fujishiro Y, Takahashi K, Sato T. Preparation and compressive strength of alpha-tricalcium phosphate/gelatin gel composite cement. *J Biomed Mater Res* 2001;54(4):525-30.
59. Miyazaki K, Horibe T, Antonucci JM, Takagi S, Chow LC. Polymeric calcium phosphate cements: analysis of reaction products and properties. *Dent Mater* 1993;9(1):41-5.
60. Miyazaki K, Horibe T, Antonucci JM, Takagi S, Chow LC. Polymeric calcium phosphate cements: setting reaction modifiers. *Dent Mater* 1993; 9(1):46-50.
61. dos Santos LA, De Oliveria LC, Rigo EC, Carrodeguas RG, Boschi AO, De Arruda AC. Influence of polymeric additives on the mechanical properties of alpha-tricalcium phosphate cement. *Bone* 1999;25(2 Suppl):99S- 102S.
62. Zhang Y, Zhang M. Calcium phosphate/chitosan composite scaffolds for controlled in vitro antibiotic drug release. *J Biomed Mater Res* 2002; 62(3):378-86.
63. Zhang Y, Xu HH. Effects of synergistic reinforcement and absorbable fiber strength on hydroxyapatite bone cement. *J Biomed Mater Res A* 2005;75(4):832-40.
64. Tajima S, Nishimoto N, Kishi Y, Matsuya S, Ishikawa K. Effects of added sodium alginate on mechanical strength of apatite cement. *Dent Mater J* 2004;23(3):329-34.
65. Ishikawa K, Miyamoto Y, Takechi M, Toh T, Kon M, Nagayama M, et al. Non-decay type fast-setting calcium phosphate cement: hydroxyapatite putty containing an increased amount of sodium alginate. *J Biomed Mater Res* 1997;36(3):393-9.
66. Bigi A, Bracci B, Panzavolta S. Effect of added gelatin on the properties of calcium phosphate cement. *Biomaterials* 2004;25(14): 2893-9.
67. Chiang T, Ho C, Chen DC, Lai M, Ding S. Physicochemical properties and biocompatibility of chitosan oligosaccharide/gelatin/calcium phosphate hybrid cements. *Mater Chem Phys* 2010;120(2-3):282-8.
68. Shie MY, Chen DC, Wang CY, Chiang TY, Ding SJ. Immersion behavior of gelatin-containing calcium phosphate cement. *Acta Biomater* 2008;4(3):646-55.
69. Bohner M, Lemaître J, Merkle HP, Gander B. Control of gentamicin release from a calcium phosphate cement by admixed poly(acrylic acid). *J Pharm Sci* 2000;89(10):1262-70.
70. Niikura T, Tsujimoto K, Yoshiya S, Tadokoro K, Kurosaka M, Shiba R. Vancomycin-impregnated calcium phosphate cement for methicillin-resistant *Staphylococcus aureus* femoral osteomyelitis. *Orthopedics* 2007;30 (4):320-1.
71. Ruhe PQ, Hedberg EL, Padron NT, Spauwen PH, Jansen JA, Mikos AG. rhBMP-2 release from injectable poly(DL-lactic-co-glycolic acid)/ calcium-phosphate cement composites. *J Bone Joint Surg Am* 2003;85-A(Suppl 3):75-81.

72. Girod Fullana S, Ternet H, Freche M, Lacout JL, Rodriguez F. Controlled release properties and final macroporosity of a pectin microspheres-calcium phosphate composite bone cement. *Acta Biomater* 2010;6(6):2294-300.
73. van de Belt H, Neut D, Uges DR, Schenk W, van Horn JR, van der Mei HC, Busscher HJ. Surface roughness, porosity and wettability of gentamicin-loaded bone cements and their antibiotic release. *Biomaterials* 2000;21(19):1981-7.
74. Takechi M, Miyamoto Y, Ishikawa K, Nagayama M, Kon M, Asaoka K, et al. Effects of added antibiotics on the basic properties of anti-washout-type fast-setting calcium phosphate cement. *J Biomed Mater Res* 1998;39(2):308-16.
75. Ratier A, Freche M, Lacout JL, Rodriguez F. Behaviour of an injectable calcium phosphate cement with added tetracycline. *Int J Pharm* 2004;274(1-2):261-8.
76. Huang Y, Liu CS, Shao HF, Liu ZJ. Study on the applied properties of tobramycin-loaded calcium phosphate cement. *Key Eng Mater* 2000;192(1):853-60.
77. Ginebra MP, Rilliard A, Fernández E, Elvira C, San Román J, Planell JA. Mechanical and rheological improvement of a calcium phosphate cement by the addition of a polymeric drug. *J Biomed Mater Res* 2001;57(1):113-8.
78. Espanol M, Perez RA, Montufar EB, Marichal C, Sacco A, Ginebra MP. Intrinsic porosity of calcium phosphate cements and its significance for drug delivery and tissue engineering applications. *Acta Biomater* 2009;5(7):2752-62.
79. Hesarakı S, Nemati R. Cephalixin-loaded injectable macroporous calcium phosphate bone cement. *J Biomed Mater Res B Appl Biomater* 2009;89(2):342-52.
80. Ratier A, Gibson IR, Best SM, Freche M, Lacout JL, Rodriguez F. Setting characteristics and mechanical behaviour of a calcium phosphate bone cement containing tetracycline. *Biomaterials* 2001;22(9):897-901.
81. Bohner M, Lemaître J, Van Landuyt P, Zambelli PY, Merkle HP, Gander B. Gentamicin-loaded hydraulic calcium phosphate bone cement as antibiotic delivery system. *J Pharm Sci* 1997;86(5):565-72.
82. Gbureck U, Vorndran E, Barralet JE. Modeling vancomycin release kinetics from microporous calcium phosphate ceramics comparing static and dynamic immersion conditions. *Acta Biomater* 2008;4(5):1480-6.
83. Alkhraisat MH, Rueda C, Cabrejos-Azama J, Lucas-Aparicio J, Mariño FT, Torres Garcia-Denche J, et al. Loading and release of doxycycline hyclate from strontium-substituted calcium phosphate cement. *Acta Biomater* 2010;6(4):1522-8.
84. Jacobs AM, Seifert AM, Kirisits TJ, Protzel HR. Use of antibiotic-loaded bone cement in the management of common infections of the foot and ankle. *Clin Podiatr Med Surg* 1990;7(3):523-44.
85. Youngman JR, Ridgway GL, Haddad FS. Antibiotic-loaded cement in revision joint replacement. *Hosp Med* 2003;64(10):613-6.
86. Diefenbeck M, Mückley T, Hofmann GO. Prophylaxis and treatment of implant-related infections by local application of antibiotics. *Injury* 2006;37(Suppl 2):S95-104.
87. Parvizi J, Saleh KJ, Ragland PS, Pour AE, Mont MA. Efficacy of antibiotic-impregnated cement in total hip replacement. *Acta Orthop* 2008;79(3):335-41.
88. van de Belt H, Neut D, Schenk W, van Horn JR, van Der Mei HC, Busscher HJ. Staphylococcus aureus biofilm formation on different gentamicin-loaded polymethylmethacrylate bone cements. *Biomaterials* 2001;22(12):1607-11.
89. Joseph TN, Chen AL, Di Cesare PE. Use of antibiotic-impregnated cement in total joint arthroplasty. *J Am Acad Orthop Surg* 2003;11(1):38-47.
90. Neut D, Hendriks JG, van Horn JR, van der Mei HC, Busscher HJ. Pseudomonas aeruginosa biofilm formation and slime excretion on antibiotic-loaded bone cement. *Acta Orthop* 2005;76(1):109-14.
91. Peter B, Gauthier O, Laïb S, Bujoli B, Guicheux J, Janvier P, et al. Local delivery of bisphosphonate from coated orthopedic implants increases implants mechanical stability in osteoporotic rats. *J Biomed Mater Res A* 2006;76(1):133-43.
92. Stadelmann VA, Gauthier O, Terrier A, Bouler JM, Pioletti DP. Implants delivering bisphosphonate locally increase periprosthetic bone density in an osteoporotic sheep model. A pilot study. *Eur Cell Mater* 2008;16:10-6.
93. Fukunaga M. [New development in bisphosphonate treatment. Review of effect on bone metabolism by minodronic acid in primary osteoporosis]. *Clin Calcium* 2009;19(1):63-73.
94. Yu T, Ye J, Gao C, Yu L, Wang Y. Synthesis and drug delivery property of calcium phosphate cement with special crystal morphology. *J Am Ceram Soc* 2010;93(5):1241-4.
95. Stallmann HP, Faber C, Bronckers AL, Nieuw Amerongen AV, Wuisman PI. In vitro gentamicin release from commercially available calcium-phosphate bone substitutes influence of carrier type on duration of the release profile. *BMC Musculoskelet Disord* 2006;7:18.
96. Doadrio JC, Arcos D, Cabañas MV, Vallet-Regí M. Calcium sulphate-based cements containing cephalixin. *Biomaterials* 2004;25(13):2629-35.
97. Yang Z, Han J, Li J, Li X, Li Z, Li S. Incorporation of methotrexate in calcium phosphate cement: behavior and release in vitro and in vivo. *Orthopedics* 2009;32(1):27.
98. Tanzawa Y, Tsuchiya H, Shirai T, Nishida H, Hayashi K, Takeuchi A, et al. Potentiation of the antitumor effect of calcium phosphate cement containing anticancer drug and caffeine on rat osteosarcoma. *J Orthop Sci* 2011;16(1):77-84.
99. Lopez-Heredia MA, Kamphuis GJ, Thüne PC, Öner FC, Jansen JA, Walboomers XF. An injectable calcium phosphate cement for the local delivery of paclitaxel to bone. *Biomaterials* 2011;32(23):5411-6.
100. Weir MD, Xu HH. Osteoblastic induction on calcium phosphate cement-chitosan constructs for bone tissue engineering. *J Biomed Mater Res A* 2010;94(1):223-33.
101. Ikawa N, Kimura T, Oumi Y, Sano T. Amino acid containing amorphous calcium phosphates and the rapid transformation into apatite. *J Mater Chem* 2009;19(28):4906-13.
102. Stallmann HP, de Roo R, Faber C, Amerongen AV, Wuisman PI. In vivo release of the antimicrobial peptide hLF1-11 from calcium phosphate cement. *J Orthop Res* 2008;26(4):531-8.
103. Stallmann HP, Faber C, Bronckers AL, Nieuw Amerongen AV, Wuisman PI. Osteomyelitis prevention in rabbits using antimicrobial peptide hLF1-11 or gentamicin-containing calcium phosphate cement. *J Antimicrob Chemother* 2004;54(2):472-6.
104. Baradari H, Damia C, Dtreih-Colas M, Champion E, Chulia D, Viana M. β -TCP porous pellets as an orthopaedic drug delivery system: ibuprofen/carrier physicochemical interactions. *Sci Technol Adv Mater* 2011;12(5):055008. doi:10.1088/1468-6996/12/5/055008
105. Jindong Z, Hai T, Junchao G, Bo W, Li B, Qiang WB. Evaluation of a novel osteoporotic drug delivery system in vitro: alendronate-loaded calcium phosphate cement. *Orthopedics* 2010;33(8). doi:10.3928/01477447-20100625-15.
106. Panzavolta S, Torricelli P, Bracci B, Fini M, Bigi A. Functionalization of biomimetic calcium phosphate bone cements with alendronate. *J Inorg Biochem* 2010;104(10):1099-106.
107. Weir MD, Xu HH. Human bone marrow stem cell-encapsulating calcium phosphate scaffolds for bone repair. *Acta Biomater* 2010;6(10):4118-26.
108. Seeherman H, Li R, Wozney J. A review of pre-clinical program development for evaluating injectable carriers for osteogenic factors. *J Bone Joint Surg Am* 2003;85-A (Suppl 3):96-108.
109. Liu WC, Wong CT, Fong MK, Cheung WS, Kao RY, Luk KD, et al. Gentamicin-loaded strontium-containing hydroxyapatite bioactive bone cement—an efficient bioactive antibiotic drug delivery system. *J Biomed Mater Res B Appl Biomater* 2010;95(2):397-406.
110. Lee GS, Park JH, Shin US, Kim HW. Direct deposited porous scaffolds of calcium phosphate cement with alginate for drug delivery and bone tissue engineering. *Acta Biomater* 2011;7(8):3178-86.
111. Zou Q, Li Y, Zhang L, Zuo Y, Li J, Li J. Antibiotic delivery system using nano-hydroxyapatite/chitosan bone cement consisting of berberine. *J Biomed Mater Res A* 2009;89(4):1108-17.
112. Rabiee SM. Development of hydroxyapatite bone cement for controlled drug release via tetracycline hydrochloride. *Bull Mater Sci* 2013;36(1):171-4.
113. Sörensen TC, Arnoldi J, Procter P, Beigel C, Jönsson A, Lennerås M, et al. Locally enhanced early bone formation of zoledronic acid incorporated into a bone cement plug in vivo. *J Pharm Pharmacol* 2013;65(2):201-12.