

Catalase-Peroxidase Gene (Kat G) Deletion in Isoniazid Resistant Strains of *Mycobacterium Tuberculosis*

İZONİAZİD DİRENÇLİ *M. TUBERCULOZİS* SUŞLARINDA KATALAZ PEROKSİDAZ GEN (KAT G) DELEZYONU

Figen ATALAY, MD,^a Nejat AKAR, MD,^b Dilek ERNAM TURGUT, MD,^a Derya AYSEV, MD,^c Pınar ERGÜN, MD,^a Yurdanur ERDOĞAN, MD,^a

^aAtatürk Chest Diseases and Chest Surgery Center,

^bDepartment of Pediatric Molecular Pathology, ^cDepartment of Microbiology, Ankara University School of Medicine, ANKARA

Abstract

Objective: A recent study showed that some isoniazid-resistant *Mycobacterium tuberculosis* strains have a complete deletion of the gene (Kat G) encoding the catalase-peroxidase enzyme. We wished to examine what proportion of clinical isolates of isoniazid-resistant *M. tuberculosis* exhibit Kat G deletion in our Country.

Material and Methods: The relation between Kat G deletion and isoniazid-resistance was examined in 10 isoniazid-resistant and 26 isoniazid-sensitive, for a total of 36 isolates, using polymerase chain reaction (PCR) technique.

Results: Kat G deletion was observed in 70% of isoniazid-resistant and 7.7% isoniazid-sensitive isolates ($p < 0.05$, χ^2). The Kat G deletion ratio in isoniazid-resistant isolates were observed to be higher than in isoniazid-sensitive isolates [odds ratio, 28, (95% CI, 3.87 to 202.26)]. The specificity and sensitivity of the PCR technique, which revealed both Kat G deletion and isoniazid-resistance, were 92.3% and 70% , respectively.

Conclusion: The deletion of the Kat G gene may play an important role in resistance to isoniazid in most *M. tuberculosis* isolates in our Country.

Key Words: Catalase-Peroxidase Gene (KatG), isoniazid resistance, MDR-TB, mycobacterium tuberculosis

T Klin J Med Sci 2004, 24:243-246

Özet

Amaç: Son zamanlarda yayınlanan çalışmalar isoniazid dirençli *M. tuberculosis* suşlarında katalaz-peroksidazı kodlayan KatG geninde total delesyonu göstermiştir. Bizde çalışmamızda isoniazid dirençli *M. tuberculosis* izolatlarında Kat G delesyonunun ülkemizdeki sıklığını araştırmayı amaçladık.

Gereç ve Yöntemler: Kat G delesyonu ile isoniazid direnci arasındaki ilişkiyi, PCR tekniği kullanarak 10 isoniazid dirençli ve 26 isoniazid hassas toplam 36 izolat üzerinde inceledik.

Bulgular: İsoniazid dirençli suşlarda % 70, isoniazid hassas suşlarda %7,7 oranında Kat G delesyonu saptandı ($p < 0,05$, χ^2). İsoniazid dirençli suşlarda Kat G delesyon oranı isoniazid hassas suşlara göre daha yüksek saptandı. Kat G delesyonu ile isoniazid direncini gösteren PCR tekniğinin spesifitesi %92,3 ve sensitivitesi %70 olarak bulundu.

Sonuç: Bu bulgular ülkemizde isoniazid dirençli çoğu *M. tuberculosis* izolatlarında direnç gelişiminde Kat G gen delesyonunun önemi vurgulamaktadır.

Anahtar Kelimeler: Katalaz -Peroksidaz Gen (KatG), isoniazid direnci, MDR-TB, m. tuberculosis

M. tuberculosis is usually treated with only a limited number of antimicrobial agents, the most important ones being rifampin, isoniazid,

streptomycin, and ethambutol.

Because of bactericidal effect and low cost, isoniazid is the most important drug used for the treatment of tuberculosis.¹ But the mechanism of its action and the development of resistance to the drug by *M. tuberculosis* has not been definitely established.² Recent advances in molecular biology have allowed identification of the genetic loci and biological mechanisms of resistance to various drugs. Previous studies have described resistance-

Geliş Tarihi/Received: 17.03.2003

Kabul Tarihi/Accepted: 27.05.2004

Yazışma Adresi/Correspondence: Dr.Figen ATALAY
Kıbrıs Sok. 17/4 06690 Aşağı Ayrancı, ANKARA,
figendr@hotmail.com

Copyright © 2004 by Türkiye Klinikleri

T Klin J Med Sci 2004, 24

243

associated mutations in *katG*, *inhA*, *kasA*, *ndh*, and the *oxy R- ahpC* intergenic region.³⁻⁸ It is proposed that INH enters *M. tuberculosis* as a prodrug by passive diffusion and is activated by catalase-peroxidase, encoded by *katG*. Mycolic acid synthesis is the primary pathway inhibited by the action of isoniazid.⁹ Two enzymes involved in the biosynthesis of mycolic acids have been suggested to be the targets of Kat G-activated isoniazid: the NADH-dependent enoyl-acyl carrier protein reductase (*inhA*) and β - ketoacyl acyl carrier protein synthase (*KasA*). Certain promotor mutations of alkylhydroperoxidase reductase, encoded by *ahpC*, in INH- resistant isolates results in overexpression of *ahpC* as compensatory mechanism for the loss of catalase activity due to *KatG* mutations.¹⁰ Recently, missense mutations were identified in *ndh*, a gene encoding NADH dehydrogenase, which is essential respiratory chain enzyme that regulates the NADH/NAD⁺ ratio in cells.⁷ Molecular techniques can detect drug resistance in *M. tuberculosis*.

We wished to examine the proportion of Kat G deletion, which we think the important molecular mechanism for isoniazid resistant strains of *M. tuberculosis* in Turkey. We examined the frequency of deletion in the *kat G* gene in isoniazid sensitive and resistant *M. tuberculosis* strains by using a PCR procedure for the first time in Turkey.

Material and Methods

Bacterial isolates: Isoniazid resistant and drug susceptible 36 *M. tuberculosis* strains isolated in randomly selected months were obtained from the Clinical Microbiology Laboratory in our hospital. All strains used in this study were grown on Lowenstein-Jensen media. Susceptibility testing of isolates was performed by using the proportion count method. Isoniazid was tested at 0.2 μ gr/ml. All isolated strains were transported in Lowenstein-Jensen culture tubes to Departments Microbiology and Pediatric Molecular Pathology in an other University.

PCR Procedure: The investigators doing the PCR procedure were blinded to the drug

susceptibility information of the isolates. *M. tuberculosis* DNA was extracted by boiling a loopful of the bacterial growth from Lowenstein-Jensen culture slants in 100 μ L of distilled water. These samples were put into sterile tubes which contains 5 ml distilled water and glass bead. Afterwards bacteria suspensions were obtained by shaking tubes in vortex. Samples were then analyzed by PCR. The PCR reaction mixture contained 200 μ M dNTP (Pharmacia Biotechnology, Piscataway, NJ), 50mM TRIS-HCl (pH 8,8), 50mM KCl, 2,5mM MgCl₂, 0,1% Triton X-100, and 0,25 units of Taq polymerase (Perkin-Elmer Cetus, Norwalk, CT). The sequences of the oligonucleotide primers (TB 1 and TB 2) used to amplify the *M. tuberculosis* catalase gene segment are as follows:

TB 1F.5'- GAG ATC GAG CTG CTG GAG GAG CC- 3'

TB 2R.5'- AGC TGC AGC CCA AAG GTG TT -3'

The sequences of oligonucleotide primers (Kat1 and Kat2) used to amplify the *M.tuberculosis* catalase gene segment are follows (Figure 1):

Kat 1F.5'-GCG ATCA CAT CAT CCG

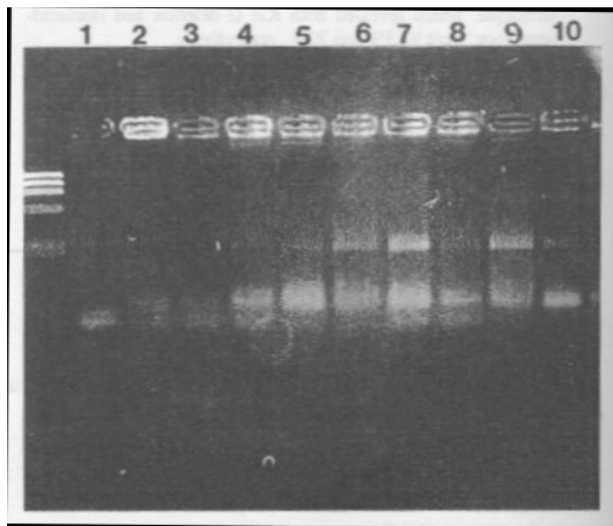


Figure 1. Analysis of KatG sequences by polymerase chain reaction (PCR)

Position of 282-bp Kat G- amplified DNA product indicated by arrow. Lanes 2- 4 strains showing absence of Kat G DNA sequences, Lanes 5- 11 strains showing presence of KatG DNA sequences

TGAT CACA-3'

Kat-2R.5'-GTCA GGCG TCA AGTC GAC
TG-3'

282-bp KatG amplified DNA products were controlled by using 2% agarose gel. All samples that yielded a negative reaction were tested at least twice and were also subjected to additional reactions with other sets of primers known to amplify other regions of *M. tuberculosis* DNA.

The PCR products were purified according to the manufacturer's instructions by E.Z.N.A. Cycle-Pure Kit (Omega Bio-tek, Doraville-USA) prior to sequencing.

Results

Thirty-six (10 isoniazid-resistant and 26 isoniazid-sensitive) isolates of *M. tuberculosis* were obtained. DNA was extracted from all isolates and we confirmed that all of them were *M. tuberculosis*. The catalase primers amplified a 282-bp DNA segment of 3 (30%) of isoniazid-resistant and 24 (92.3%) of isoniazid-sensitive isolates ($p < 0.05$, χ^2 test) (Table 1). The catalase primers repeatedly failed to amplify DNA extracted from 7 (70%) isoniazid-resistant and 2 (7.7%) isoniazid-sensitive isolates. Kat G deletion ratio in isoniazid resistant isolates were observed higher than isoniazid sensitive isolates [odds ratio, 28, (95% CI, 3.87 to 202.26)]. For katG deletion, the specificity, sensitivity, positive and negative predictive values of the assay compared with conventional methods were 92.3%, 70%, 77.8% and 88.8%, respectively.

Discussion

The incidence of tuberculosis is increasing in many countries, and control of the disease is

further threatened by the emergence of drug resistance. Recent advances in molecular biology have allowed identification of the genetic loci and biological mechanisms of resistance to various drugs.^{2-4,11-18} For the first time, 1992, the relationship between isoniazid-resistance and loss of catalase activity were explained by the identification of the katG gene, encoding the catalase-peroxidase enzyme of *M. tuberculosis* which is indispensable for isoniazid susceptibility. Zhang et al. found that total deletion of the katG gene occurs in ~20% of isoniazid-resistant isolates.³ Furthermore, they showed that 2 of 3 resistant strains of *M. tuberculosis* strains completely lack of the catalase gene. They concluded that in a subset of isoniazid resistant isolates, the lack of catalase activity is due to the complete loss of the Kat G gene.¹³ Initially, resistance was believed to be caused by complete deletion of the Kat G gene, but with the use of PCR and PCR-SSCP, it was shown that both deletion and mutation of the Kat G gene could lead to resistance.¹⁹

Stoeckle et al found that 31 (76%) of 41 isoniazid-resistant isolates in New York City contained Kat G sequences. These findings suggest that the potential diagnostic application of Kat G sequence as a marker for isoniazid resistance would yield a test of low sensitivity and specificity.²⁰

In the present study, we found that 3 (30%) of isoniazid resistant and 24 (92.3%) of isoniazid sensitive isolates contained Kat G sequences ($p < 0.05$, χ^2). A significantly higher katG deletion was found in isoniazid-resistant strains compared with isoniazid-sensitive strains.

Table 1. Evaluation of Kat G deletion in isoniazid resistant and sensitive patients

	Isoniazid- resistant strains of <i>M. tuberculosis</i> , n (%)	isoniazid-sensitive strains of <i>M. tuberculosis</i> , n (%)	Total, n
Kat G deletion positive	7 (70)	2 (7.7)	9
Kat G deletion negative	3 (30)	24 (92.3)	27
Total, n	10	26	36

χ^2 , $p < 0.05$

When compared our findings with Stoeckle et al., we found similar Kat G deletion rates in isoniazid sensitive strains but significant was found in isoniazid-resistant strains (24% versus 70%). This could be explained by the difference in geographical origins. In the study of Abal et al.²¹ from Kuwait, the mutation S315T was found in 46 (69%) of the 67 resistant strains. The prevalence of this mutation was highest (80%) in isolates recovered from patients of South Asian origin and lowest in isolates from patients of Middle Eastern origin (44%). These results point to a varying prevalence of the mutation within the Kat G gene in clinical *M. tuberculosis* isolates recovered from patients of different ethnic groups within the same country.

The specificity and sensitivity of PCR technique, which reveals katG deletion and isoniazid-resistance, is 92.3% and 70%, respectively. These findings support the importance of katG deletion in the development of isoniazid-resistance in Turkey.

Finally, resistance to isoniazid in most *M. tuberculosis* isolates is due to the mutations on KatG gene in Turkey. But there is a further need to identify KatG variations in *M. tuberculosis* isolates.

REFERENCES

- Iseman MD. Clinician's Guide to Tuberculosis. 1st ed. Philadelphia: Lippincott Williams & Wilkins Co; 2000. p.274-323.
- Telenti A. Genetics of drug resistance in tuberculosis. Clin Chest Med 1997; 18: 55-54.
- Zhang Y, Heym B, Allen B, Young D and Cole S. The catalase- peroxidase gene and isoniazid resistance of *Mycobacterium tuberculosis*. Nature 1992; 358: 591-3.
- Heym B, Zhang Y, Poulet S, et al. Characterization of the katG gene encoding a catalase-peroxidase required for the isoniazid susceptibility of *Mycobacterium tuberculosis*. J Bact 1993; 175: 4255-59.
- Mdluli K, Slayden RA, Zhu Y, et al. Inhibition of a *Mycobacterium tuberculosis* β -ketoacyl ACP synthase by isoniazid. Science 1998;280: 1607-10.
- Musser JM, Kapur V, Williams DL, et al. Characterization of the catalase-peroxidase gene (Kat G) and inhA locus in isoniazid-resistant and -susceptible strains of *Mycobacterium tuberculosis* by automated DNA sequencing: restricted array of mutations associated with drug resistance. J Infect Dis 1996; 173:196-2.
- Lee ASG, Teo ASM, and Wong SY. Novel mutations in *ndh* in isoniazid-resistant *Mycobacterium tuberculosis* isolates. Antimicrob Agents Chemother 2001; 45: 2157-9.
- Ramaswamy SV, Reich R, Dou SJ, et al. Single nucleotide polymorphisms in genes associated with isoniazid resistance in *Mycobacterium tuberculosis*. Antimicrob Agents Chemother 2003; 47: 1241-49.
- Winder FG, and Collins PB. Inhibition by isoniazid of synthesis of mycolic acids in *Mycobacterium tuberculosis*. J Gen Microbiol 1970 ; 63: 41-8.
- Lee ASG, Teo ASM, and Wong SY. Novel mutations in *ndh* in isoniazid-resistant *Mycobacterium tuberculosis* isolates. Antimicrob Agents Chemother 2001; 45: 2157-59.
- Heym B, Alzari PM, Honore N, Cole S.T. Missense mutations in catalase-peroxidase gene katG are associated with isoniazid resistance in *Mycobacterium tuberculosis*. Mol Microbiol 1995; 15: 235-5.
- Rastogi N, David HL. Mode of Action of Antituberculous drugs and mechanism of drug resistance in *Mycobacterium tuberculosis*. Res Microbiol 1993; 144: 133-43.
- Zhang Y. Genetic Basis of Isoniazid resistance of *Mycobacterium tuberculosis*. Res Microbiol 1993; 144: 143-9.
- Telenti A, Imboden P, Marchesi F, et al. Direct, automated detection of rifampin-resistant *Mycobacterium tuberculosis* by polymerase chain reaction and single-strand conformation polymorphism analysis. Anti Microb Agents Chemother 1993;37:2054-8.
- Telenti A, Imboden P, Marchesi D, et al. Detection of rifampisin-resistant mutations in *Mycobacterium tuberculosis*. Lancet 1993; 341: 647-50.
- Heym B, Honoré N, Truffot-Pernot C, et al. Implications of multidrug resistance for the future of short-course chemotherapy of tuberculosis: a molecular study. Lancet 1994; 344: 293-8.
- Nachamkin I, Kang C, Weinstein PM. Detection of resistance to isoniazid, rifampin, and streptomycin in clinical isolates of *Mycobacterium tuberculosis* by Molecular Methods. Clinical Infect Dis 1997; 724: 894-900.
- Heym B, Saint-Joanis B, Cole ST. The molecular basis of isoniazid resistance in *Mycobacterium tuberculosis*. Tubercle and Lung Disease 1997; 79(4): 267-71.
- Fluit AC, Visser MR, and Schmitz F. Molecular detection of antimicrobial resistance. Clinical Microbiology Reviews 2001; 14 (4): 836- 71
- Stoeckle MY, Guan L, Riegler N, et al. Catalase-Peroxidase gene sequences in isoniazid-sensitive and -resistant strains of *Mycobacterium tuberculosis* from New York City. J Infect Dis 1993; 168: 1063-65.
- Abal AT, Ahmad S, and Mokaddas E. Variations in the occurrence of the S315T mutation within Kat G gene in isoniazid-resistant clinical *Mycobacterium tuberculosis* isolates from Kuwait. Microb Drug Resist 2002; 8 (2): 99-105.